

A METABOLIC MECHANISM FOR DIABETIC NEUROPATHY

DIVIJA DESHPANDE¹, ASA HIDMARK², THOMAS FLEMING³, PETER NAWROTH⁴

^{1,2,3,4}Department of Medicine I and Clinical Chemistry, University Hospital of Heidelberg, INF 410, 69120 Heidelberg, Germany

Keywords: diabetic neuropathy, mechanism, metabolism

Abstract: Elevated blood glucose alone cannot explain the development and progression of diabetic neuropathy (DN) and the lowering of blood glucose is insufficient in preventing and/or reversing neuropathy in patients with type 2 diabetes. Dicarbonyls, such as methylglyoxal (MG), are endogenous by-products of glycolysis, which are elevated in diabetic patients. Reactive metabolite such as MG can modify DNA as well as extra- and intracellular proteins, leading to the formation of advanced glycation endproducts (AGEs). MG can contribute to the development of DN via post-translational modification of neuronal ion channels involved in chemosensing and action potential generation in nociceptive nerve endings. Increased formation of AGEs leads to increased cellular stress, dysfunction and ultimately cell death. The interaction of AGE-modified proteins through cell surface receptors, such as RAGE, can lead to increased cellular activation and sustained inflammatory responses. An inflammatory immune response is associated with later, degenerative, stages of DN. The direct and indirect effects of dicarbonyls on nerves provides a unifying mechanism for the development and progression of DN. Preventing the accumulation of MG during diabetes and/or inhibiting of RAGE signalling may therefore provide new, more effective, therapeutic approaches for the treatment of DN.

Cuvinte cheie: neuropatie diabetică, mecanism, metabolism

Rezumat: Doar valorile crescute ale glucozei în sânge nu pot explica dezvoltarea și avansarea neuropatiei diabetice, iar scăderea valorilor glucozei în sânge nu este suficientă pentru a preveni neuropatia la pacienții cu diabet zaharat de tip 2. Dicarboxili cum ar fi metilglioxalul sunt produși endogeni secundari glicolizei, înregistrând valori crescute la pacienții diabetici. Metabolitul reactiv, de exemplu metilglioxalul poate modifica ADN-ul, precum și proteinele extracelulare și intracelulare, conducând la formarea de produși finali de glicozilare avansată. Metilglioxalul poate contribui la dezvoltarea neuropatiei diabetice prin modificarea post - translațională a canalelor ionice neuronale implicate în chemosensibilitate și generarea de acțiune potențială în terminațiile nervoase nociceptive. Formarea crescută de produși finali de glicozilare avansată duce la creșterea stresului celular, disfuncție și în cele din urmă la moartea celulei. Interacțiunea proteinelor modificate ca urmare a produșilor finali de glicozilare avansată prin receptori celulari de suprafață, cum ar fi receptorii pentru produși finali de glicozilare avansată, poate duce la creșterea activării celulare și la răspunsuri inflamatorii susținute. Un răspuns imun inflamator este asociat cu etapele ulterioare, degenerative ale neuropatiei diabetice. Efectele directe și indirecte ale dicarbonililor asupra nervilor oferă un mecanism unificator pentru dezvoltarea și avansarea neuropatiei diabetice. Prevenirea acumulării de metilglioxal în caz de diabet și / sau inhibarea semnalizării receptorilor pentru produși finali de glicozilare avansată (RAGE) poate astfel oferi abordări terapeutice noi, mai eficiente privind tratamentul neuropatiei diabetice.

Diabetes is a disease posing considerable economic burden on global health services. Approximately 50% of the diabetic patients develop diabetic neuropathy (DN) as a late complication.(1) DN is well characterized in the peripheral nervous system and has a distal predominance. Patients with DN display altered pain sensitivities such as increased pain sensitivity (*hyperalgesia*) as well as hyper-responsiveness to normally innocuous stimuli (*allodynia*) during the early stages of the diseases in the initial stages. Symptoms of sensory loss may be present throughout the course of the disease, but predominate in the later stages as a result of the degeneration of neurons and their myelin sheaths.(2) Since hyperglycemia characterizes diabetes, tight control of blood glucose continues to be the primary treatment option. However, the drawback of intensive glycemic control is increased occurrence of hypoglycaemic episodes with a risk of brain injury and death.

Moreover, normalizing blood glucose does not significantly reduce the incidence of neuropathic symptoms in type II diabetic patients and can only marginally restore the abnormal nerve functions in type I diabetic patients.(3–8) These studies indicate that the pathways in which the metabolic flux is directed or redirected are more important than the increased level of glucose itself when considering a treatment strategy for DN. This conclusion is supported by a study in which it was shown that diabetic patients are more prone to accumulate triose phosphates (glycolytic intermediate) than healthy controls, even after normalization to individual glucose concentrations.(9) In diabetic patients, the concentration of triose phosphates correlated positively with the concentration of the reactive metabolite methylglyoxal (MG), formed by non-enzymatic dephosphorylation of these intermediates. This study showed that even at normal glucose levels, diabetic patients have

¹Corresponding author: Peter P. Nawroth, Medicine I and Clinical Chemistry, University Hospital of Heidelberg, INF 410, 69120 Heidelberg, Germany, Email: peter.nawroth@med.uni-heidelberg.de, Tel: +496221 568601

Article received on 19.01.2014 and accepted for publication on 28.01.2014

ACTA MEDICA TRANSILVANICA March 2014;2(1):141-144

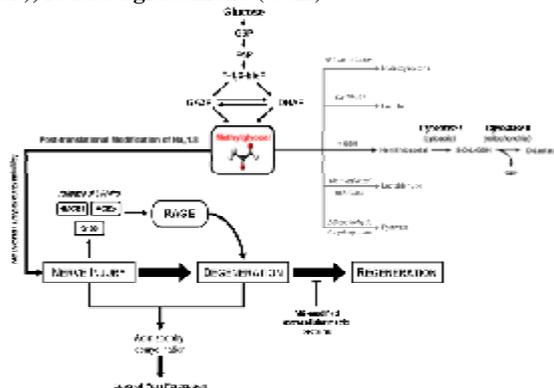
CLINICAL ASPECTS

dramatically increased concentrations of glucose-derived reactive metabolites.

MG-induced hyperexcitability: a mechanism for hyperalgesia in the early stages of DN.

Hyperglycemia will lead to an increased flux through glycolysis and increased production of MG through the accumulation of triose phosphates.(10) (figure no. 1). Neuronal tissue has high energy demands and absorbs large amounts of glucose, independent of insulin even under normoglycemic conditions, and as such, is at higher risk for the accumulation of MG. MG can be metabolized by (i) MG reductase to lactaldehyde (11); (ii) aldose reductase isozymes, such as Akr1b1, which the NADPH-dependent reduction of MG to hydroxyacetone (12); (iii) betaine aldehyde dehydrogenase, also known as Aldh7a1, which catalyzes the oxidation of MG to pyruvate (13) and (iv) 2-oxoaldehyde dehydrogenase which can also catalyze the oxidation of MG to pyruvate.(13)

Figure no. 1. The Production & Detoxification of Methylglyoxal and its role in the Diabetic Neuropathy. Glucose-6-phosphate (G6P), Fructose-1, 6-bisphosphate (F-1, 6-biP), Glyceraldehyde-3-phosphate (GA3P), Dihydroxyacetone phosphate (DHAP), Ribose-5-phosphate (R5P), reduced glutathione (GSH)



Recent research has identified that DJ1/Park7 can function to metabolize MG directly to lactic acid, independent of glutathione.(14) However, the majority of the MG produced during glycolysis is metabolized to the stable end product, D-lactate by glyoxalase 1 (Glo1) and glyoxalase 2 (Glo2) along with catalytic amounts of reduced glutathione (GSH).(15) It has been shown that within the context of diabetic neuropathy, sciatic nerves are more susceptible to MG due to a reduction in the transcription, expression and activity of Glo1, although the mechanism for this down regulation remains uncertain. Mice with peritoneal damage represent a good model to study Glo1 activity.(16) Reduction in the amount of Glo1 in peripheral nerves of diabetic mice has been correlated with increased sensitivity to mechanical pain.(17) Mouse-strains harboring multiple copies of the Glo1 gene are less prone to developing diabetic hyperalgesia and reduced of nerve fiber density, as compared to mice with a single copy of the gene.(18) However, to interpret pain measurements from genetically modified mice induced with diabetes, it is important to critically evaluate the mouse model to allow conclusions relevant for human DN.(19)

The association between reduced Glo1 activity and painful DN has also been shown in clinical studies performed on diabetic patients (20). Furthermore, it has been demonstrated that knock-down of Glo1 in *C.elegans*, a well characterized model for studying mechanisms of neuropathy, suitable because of its easily accessible nervous system, resulted in accelerated neuronal degeneration associated with aging.(21,22) Thus, importance of has not only been implicated in maintenance of

nerve function, but also neuronal integrity. MG, accumulated in the neuronal tissue during diabetes, can react irreversibly with the free amino groups of the lysine and arginine to generate advanced glycation endproducts (AGEs).(10,23) Numerous studies have been published showing increased glycation of neuronal tissue in diabetes.(24–31) Elevated MG and AGEs have been shown to modulate pain perception by modifying ion channels present in the peripheral nerves. Bierhaus et al showed that MG at concentrations found in the plasma of diabetic patients can rapidly modify arginine residues within the voltage-gated sodium channel Nav1.8, a channel protein playing an essential role in acute and chronic pain.(32) Modification of Nav1.8 by MG is associated with increased electrical excitability and facilitated firing of nociceptive neurons. MG treatment likewise causes a depolarizing shift in the resting membrane potential of cultured DRG neurons and reduces current threshold for activation of action potentials. The reduction in depolarization required to reach the voltage threshold, observed in the presence of MG, is independent of the Nav1.8 MG-modification. The MG-induced depolarization could contribute to increased neuronal excitability by facilitating the activation of Nav1.8 that normally has a relative high voltage threshold. Comprehensive findings have also been reported for the transient receptor potential channel subfamily A, member 1 which serves as a almost universal chemosensor in nociceptors (TRPA1).(33,34) Thus, a metabolic change resulting in excessive accumulation and reduced detoxification of MG can explain nerve dysfunction during early diabetes. However, the effect of MG and the consequences of MG-induced hyperexcitability may also provide the conditions by which the characteristic symptoms of the late stage of DN, through induction of neuronal damage via activation of an immune response.

Does MG-induced hyperexcitability induce inflammation-driven neuronal injury?

Continued metabolic insults during diabetes provoke an inflammatory response in the neuronal tissue.(35–38) In the peripheral nerves, endoneurial resident macrophages and Schwann cells, which are the nerve-resident phagocytic cells, can produce proinflammatory cytokines such as IL-1 β , IL-6 and TNF- α upon activation (39,37,40). The inflammatory response is exacerbated by macrophages and lymphocytes infiltrating into the peripheral nerve tissue.(41–43) Proinflammatory milieu in central nervous system is mediated by microglial cells during diabetes.(44–46) AGEs formed during diabetes bind to RAGE (Receptor for Advanced Glycation End Products) and trigger of signalling pathways that result in sustained activation of the proinflammatory transcription factor NF- κ B.(37,47,48) Expression of RAGE on hematopoietic cells has recently been shown to be sufficient for preventing regeneration after injury during diabetes (49). Using mouse models of neuropathic pain, several studies have linked the proinflammatory cytokines to hyperalgesia.(50) The nervous tissue is subject to ischemic injury during diabetes. Schwann cells and macrophages initiate nerve fibre loss distal to the site of injury. This process is termed Wallerian degeneration. Schwann cells also begin demyelination of the long axons of the peripheral nerves.(51–53) Under hyperglycaemic conditions, Schwann cells have been reported to undergo apoptosis.(54) MG-induced activation of p38 MAPK pathway has shown to be responsible for Schwann cell death.(55) Under normal conditions, the immune response involves an anti-inflammatory phase follows the proinflammatory phase, wherein the regeneration of the damaged nerve occurs. The transparent embryos zebrafish can be used to study nerve regeneration during diabetes, permitting neurite growth to be imaged.(56) In diabetes, the nerve

CLINICAL ASPECTS

regeneration process, which is orchestrated by Schwann cells, is greatly impaired. Hyperglycemia suppresses Schwann cell proliferation, which is prerequisite for nerve regeneration and also diminishes neurite regeneration length. MG modification has been implicated in failure of nerve regeneration and collateral sprouting.(57,58) Thus, in later stages of diabetes, progressive loss of neuronal integrity, myelination in combination with compromised nerve regeneration accounts for loss of sensory perception.

Conclusion:

An imbalance between production and detoxification of reactive metabolites provides a unifying mechanism for the development and progression of DN. The early symptoms of the disease are mediated by structural and functional modifications of membrane-bound ion channel proteins, whilst the later stages are mediated by either direct actions of MG on intracellular targets or, indirectly, through modulation of the immune response by RAGE activation. Whether the altered metabolism in the nerves induces and shapes a local inflammation during DN, or if systemic metabolic effects of diabetes on the immune system determines the inflammation in the nerve remains a question.

Acknowledgements

This work was supported by grants from the Deutsche Forschungsgemeinschaft (BI-1281/3-1 & NA 138 /7-1) and the Dietmar-Hopp-Stiftung.

REFERENCES

1. Danaei G, Finucane MM, Lu Y, Singh GM, Cowan MJ, Paciorek CJ, et al. National, regional, and global trends in fasting plasma glucose and diabetes prevalence since 1980: systematic analysis of health examination surveys and epidemiological studies with 370 country-years and 2.7 million participants. *Lancet*. 2011 Jul 2;378(9785):31–40.
2. Callaghan BC, Cheng HT, Stables CL, Smith AL, Feldman EL. Diabetic neuropathy: clinical manifestations and current treatments. *Lancet Neurol*. 2012 Jun;11(6):521–34.
3. Callaghan BC, Little AA, Feldman EL, Hughes RAC. Enhanced glucose control for preventing and treating diabetic neuropathy. *Cochrane Database Syst Rev*. 2012;6:CD007543.
4. Ismail-Beigi F, Craven T, Banerji MA, Basile J, Calles J, Cohen RM, et al. Effect of intensive treatment of hyperglycaemia on microvascular outcomes in type 2 diabetes: an analysis of the ACCORD randomised trial. *Lancet*. 2010 Aug 7;376(9739):419–30.
5. Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). UK Prospective Diabetes Study (UKPDS) Group. *Lancet*. 1998 Sep 12;352(9131):837–53.
6. Duckworth W, Abraira C, Moritz T, Reda D, Emanuele N, Reaven PD, et al. Glucose control and vascular complications in veterans with type 2 diabetes. *N Engl J Med*. 2009 Jan 8;360(2):129–39.
7. Bongaerts BWC, Rathmann W, Kowall B, Herder C, Stöckl D, Meisinger C, et al. Postchallenge hyperglycemia is positively associated with diabetic polyneuropathy: the KORA F4 study. *Diabetes Care*. 2012 Sep;35(9):1891–3.
8. Lu B, Hu J, Wen J, Zhang Z, Zhou L, Li Y, et al. Determination of peripheral neuropathy prevalence and associated factors in Chinese subjects with diabetes and pre-diabetes - ShangHai Diabetic neuropathy Epidemiology and Molecular Genetics Study (SH-DREAMS). *PloS One*. 2013;8(4):e61053.
9. Fleming T, Cuny J, Nawroth G, Djuric Z, Humpert PM, Zeier M, et al. Is diabetes an acquired disorder of reactive glucose metabolites and their intermediates? *Diabetologia*. 2012 Apr;55(4):1151–5.
10. Thornalley PJ, Jahan I, Ng R. Suppression of the accumulation of triosephosphates and increased formation of methylglyoxal in human red blood cells during hyperglycaemia by thiamine in vitro. *J Biochem (Tokyo)*. 2001 Apr;129(4):543–9.
11. Ray M, Ray S. Purification and partial characterization of a methylglyoxal reductase from goat liver. *Biochim Biophys Acta*. 1984 Nov 6;802(1):119–27.
12. Vander Jagt DL, Robinson B, Taylor KK, Hunsaker LA. Reduction of trioses by NADPH-dependent aldo-keto reductases. Aldose reductase, methylglyoxal, and diabetic complications. *J Biol Chem*. 1992 Mar 5;267(7):4364–9.
13. Vander Jagt DL, Hunsaker LA. Methylglyoxal metabolism and diabetic complications: roles of aldose reductase, glyoxalase-I, betaine aldehyde dehydrogenase and 2-oxoaldehyde dehydrogenase. *Chem Biol Interact*. 2003 Feb 1;143-144:341–51.
14. Lee J, Song J, Kwon K, Jang S, Kim C, Baek K, et al. Human DJ-1 and its homologs are novel glyoxalases. *Hum Mol Genet*. 2012 Jul 15;21(14):3215–25.
15. Thornalley PJ. Glyoxalase I--structure, function and a critical role in the enzymatic defence against glycation. *Biochem Soc Trans*. 2003 Dec;31(Pt 6):1343–8.
16. Müller-Krebs S, Zhang W, Kihm LP, Reiser J, Nawroth PP, Schwenger V. Glucose effects on the peritoneum: what can we learn from rodent models? *Exp Clin Endocrinol Diabetes Off J Ger Soc Endocrinol Ger Diabetes Assoc*. 2012 Apr;120(4):197–8.
17. Jack MM, Ryals JM, Wright DE. Characterisation of glyoxalase I in a streptozocin-induced mouse model of diabetes with painful and insensate neuropathy. *Diabetologia*. 2011 Aug;54(8):2174–82.
18. Jack MM, Ryals JM, Wright DE. Protection from diabetes-induced peripheral sensory neuropathy--a role for elevated glyoxalase I? *Exp Neurol*. 2012 Mar;234(1):62–9.
19. Bierhaus A, Nawroth PP. Critical evaluation of mouse models used to study pain and loss of pain perception in diabetic neuropathy. *Exp Clin Endocrinol Diabetes Off J Ger Soc Endocrinol Ger Diabetes Assoc*. 2012 Apr;120(4):188–90.
20. Skapare E, Konrade I, Liepinsh E, Strele I, Makrecka M, Bierhaus A, et al. Association of reduced glyoxalase 1 activity and painful peripheral diabetic neuropathy in type 1 and 2 diabetes mellitus patients. *J Diabetes Complications*. 2013 Jun;27(3):262–7.
21. Mendler M, Schlotterer A, Morcos M, Nawroth PP. Understanding diabetic polyneuropathy and longevity: what can we learn from the nematode *Caenorhabditis elegans*? *Exp Clin Endocrinol Diabetes Off J Ger Soc Endocrinol Ger Diabetes Assoc*. 2012 Apr;120(4):182–3.
22. Morcos M, Du X, Pfisterer F, Hutter H, Sayed AAR, Thornalley P, et al. Glyoxalase-1 prevents mitochondrial protein modification and enhances lifespan in *Caenorhabditis elegans*. *Aging Cell*. 2008 Mar;7(2):260–9.
23. Thornalley PJ. Protein and nucleotide damage by glyoxal and methylglyoxal in physiological systems--role in ageing and disease. *Drug Metabol Drug Interact*. 2008;23(1-2):125–50.
24. Vlassara H, Brownlee M, Cerami A. Nonenzymatic glycosylation of peripheral nerve protein in diabetes mellitus. *Proc Natl Acad Sci U S A*. 1981 Aug;78(8):5190–2.
25. Vlassara H, Brownlee M, Cerami A. Excessive nonenzymatic glycosylation of peripheral and central

CLINICAL ASPECTS

- nervous system myelin components in diabetic rats. *Diabetes*. 1983 Jul;32(7):670-4.
26. Williams SK, Howarth NL, Devenny JJ, Bitensky MW. Structural and functional consequences of increased tubulin glycosylation in diabetes mellitus. *Proc Natl Acad Sci U S A*. 1982 Nov;79(21):6546-50.
 27. Cullum NA, Mahon J, Stringer K, McLean WG. Glycation of rat sciatic nerve tubulin in experimental diabetes mellitus. *Diabetologia*. 1991 Jun;34(6):387-9.
 28. Sugimoto K, Nishizawa Y, Horiuchi S, Yagihashi S. Localization in human diabetic peripheral nerve of N(epsilon)-carboxymethyllysine-protein adducts, an advanced glycation endproduct. *Diabetologia*. 1997 Dec;40(12):1380-7.
 29. Ryle C, Donaghy M. Non-enzymatic glycation of peripheral nerve proteins in human diabetics. *J Neurol Sci*. 1995 Mar;129(1):62-8.
 30. Ryle C, Leow CK, Donaghy M. Nonenzymatic glycation of peripheral and central nervous system proteins in experimental diabetes mellitus. *Muscle Nerve*. 1997 May;20(5):577-84.
 31. Sensi M, Morano S, Morelli S, Castaldo P, Sagratella E, De Rossi MG, et al. Reduction of advanced glycation end-product (AGE) levels in nervous tissue proteins of diabetic Lewis rats following islet transplants is related to different durations of poor metabolic control. *Eur J Neurosci*. 1998 Sep;10(9):2768-75.
 32. Bierhaus A, Fleming T, Stoyanov S, Leffler A, Babes A, Neacsu C, et al. Methylglyoxal modification of Nav1.8 facilitates nociceptive neuron firing and causes hyperalgesia in diabetic neuropathy. *Nat Med*. 2012 Jun;18(6):926-33.
 33. Rush AM, Dib-Hajj SD, Liu S, Cummins TR, Black JA, Waxman SG. A single sodium channel mutation produces hyper- or hypoexcitability in different types of neurons. *Proc Natl Acad Sci U S A*. 2006 May 23;103(21):8245-50.
 34. Eberhardt MJ, Filipovic MR, Leffler A, De La Roche J, Kistner K, Fischer MJ, et al. Methylglyoxal activates nociceptors through TRPA1 - a possible mechanism of metabolic neuropathies. *J Biol Chem*. 2012;
 35. Vincent AM, Callaghan BC, Smith AL, Feldman EL. Diabetic neuropathy: cellular mechanisms as therapeutic targets. *Nat Rev Neurol*. 2011 Oct;7(10):573-83.
 36. Hur J, Sullivan KA, Pande M, Hong Y, Sima AAF, Jagadish HV, et al. The identification of gene expression profiles associated with progression of human diabetic neuropathy. *Brain J Neurol*. 2011 Nov;134(Pt 11):3222-35.
 37. Sbai O, Devi TS, Melone MAB, Feron F, Khrestchatsky M, Singh LP, et al. RAGE-TXNIP axis is required for S100B-promoted Schwann cell migration, fibronectin expression and cytokine secretion. *J Cell Sci*. 2010 Dec 15;123(Pt 24):4332-9.
 38. Hotamisligil GS. Inflammation and metabolic disorders. *Nature*. 2006 Dec 14;444(7121):860-7.
 39. Nukada H, McMorrnan PD, Baba M, Ogasawara S, Yagihashi S. Increased susceptibility to ischemia and macrophage activation in STZ-diabetic rat nerve. *Brain Res*. 2011 Feb 10;1373:172-82.
 40. Murwani R, Hodgkinson S, Armati P. Tumor necrosis factor alpha and interleukin-6 mRNA expression in neonatal Lewis rat Schwann cells and a neonatal rat Schwann cell line following interferon gamma stimulation. *J Neuroimmunol*. 1996 Dec;71(1-2):65-71.
 41. Machelska H. Dual peripheral actions of immune cells in neuropathic pain. *Arch Immunol Ther Exp (Warsz)*. 2011 Feb;59(1):11-24.
 42. Younger DS. Diabetic neuropathy: a clinical and neuropathological study of 107 patients. *Neurol Res Int*. 2010;2010:140379.
 43. Conti G, Stoll G, Scarpini E, Baron PL, Bianchi R, Livraghi S, et al. p75 neurotrophin receptor induction and macrophage infiltration in peripheral nerve during experimental diabetic neuropathy: possible relevance on regeneration. *Exp Neurol*. 1997 Jul;146(1):206-11.
 44. Tsuda M, Ueno H, Kataoka A, Tozaki-Saitoh H, Inoue K. Activation of dorsal horn microglia contributes to diabetes-induced tactile allodynia via extracellular signal-regulated protein kinase signaling. *Glia*. 2008 Mar;56(4):378-86.
 45. Zhuo M, Wu G, Wu L-J. Neuronal and microglial mechanisms of neuropathic pain. *Mol Brain*. 2011;4:31.
 46. Feng Y, Busch S, Gretz N, Hoffmann S, Hammes H-P. Crosstalk in the retinal neurovascular unit - lessons for the diabetic retina. *Exp Clin Endocrinol Diabetes Off J Ger Soc Endocrinol Ger Diabetes Assoc*. 2012 Apr;120(4):199-201.
 47. Mahajan N, Dhawan V. Receptor for advanced glycation end products (RAGE) in vascular and inflammatory diseases. *Int J Cardiol*. 2013 Oct 3;168(3):1788-94.
 48. Yeh CH, Sturgis L, Haidacher J, Zhang XN, Sherwood SJ, Bjercke RJ, et al. Requirement for p38 and p44/p42 mitogen-activated protein kinases in RAGE-mediated nuclear factor-kappaB transcriptional activation and cytokine secretion. *Diabetes*. 2001 Jun;50(6):1495-504.
 49. Juranek JK, Geddis MS, Song F, Zhang J, Garcia J, Rosario R, et al. RAGE deficiency improves postinjury sciatic nerve regeneration in type 1 diabetic mice. *Diabetes*. 2013 Mar;62(3):931-43.
 50. Austin PJ, Moalem-Taylor G. The neuro-immune balance in neuropathic pain: involvement of inflammatory immune cells, immune-like glial cells and cytokines. *J Neuroimmunol*. 2010 Dec 15;229(1-2):26-50.
 51. Greene DA, Sima AA, Stevens MJ, Feldman EL, Lattmer SA. Complications: neuropathy, pathogenetic considerations. *Diabetes Care*. 1992 Dec;15(12):1902-25.
 52. Feldman EL, Stevens MJ, Greene DA. Pathogenesis of diabetic neuropathy. *Clin Neurosci N Y N*. 1997;4(6):365-70.
 53. Wellen KE, Hotamisligil GS. Inflammation, stress, and diabetes. *J Clin Invest*. 2005 May;115(5):1111-9.
 54. Delaney CL, Russell JW, Cheng HL, Feldman EL. Insulin-like growth factor-I and over-expression of Bcl-xL prevent glucose-mediated apoptosis in Schwann cells. *J Neuropathol Exp Neurol*. 2001 Feb;60(2):147-60.
 55. Fukunaga M, Miyata S, Higo S, Hamada Y, Ueyama S, Kasuga M. Methylglyoxal induces apoptosis through oxidative stress-mediated activation of p38 mitogen-activated protein kinase in rat Schwann cells. *Ann N Y Acad Sci*. 2005 Jun;1043:151-7.
 56. Jörgens K, Hillebrands J-L, Hammes H-P, Kroll J. Zebrafish: a model for understanding diabetic complications. *Exp Clin Endocrinol Diabetes Off J Ger Soc Endocrinol Ger Diabetes Assoc*. 2012 Apr;120(4):186-7.
 57. Gumy LF, Bampton ETW, Tolkovsky AM. Hyperglycaemia inhibits Schwann cell proliferation and migration and restricts regeneration of axons and Schwann cells from adult murine DRG. *Mol Cell Neurosci*. 2008 Feb;37(2):298-311.
 58. Duran-Jimenez B, Dobler D, Moffatt S, Rabbani N, Streuli CH, Thornalley PJ, et al. Advanced glycation end products in extracellular matrix proteins contribute to the failure of sensory nerve regeneration in diabetes. *Diabetes*. 2009 Dec;58(12):2893-903.